

Annexure A

A1. MATERIALS

All the chemicals except wherever mentioned, are procured from Sigma Aldrich, India and are used without any further purification. Analytical graded reagents are used in all cases. Double distilled water is used throughout for the preparation of the samples. Samples of agrochemicals and their fragments (metabolites) are prepared within an isolated chamber, considering the safety issues. The description of chemicals is given in table A1.

Table A1: List of chemicals used and relevant description, molecular weight (M.wt.), and procurement.

S.No.	Chemicals	Description	M. wt.
1.	Bovine Serum Albumin (Plasma Protein)	98% purity, nuclease and protease free (<i>Hi-media laboratories Pvt. Ltd., India</i>)	66.5 KDa
2.	Human Serum Albumin	Plasma Protein 98% purity, nuclease free	66.5 KDa
3.	Trypsin	Enzymatic Protein	23.3 KDa
4.	Melittin	Peptide	2840 g/mol
5.	β -Cyclodextrin (C ₄₂ H ₇₀ O ₃₅)	Cyclomaltoheptaose (<i>Alfa Aesar, India</i>)	1134.9 g/mol
6.	Warfarin	4-hydroxy-3-(3-oxo-1-phenylbutyl)chromen-2-one	308.3 g/mol
7.	Ibuprofen	2-[4-(2-methylpropyl)phenyl] propanoic acid	206.3 g/mol
8.	Tryptophan	(2S)-2-amino-3-(1H-indol-3-yl) propanoic acid	204.2 g/mol
9.	Oleic acid	Fatty acid (<i>Alfa Aesar, India</i>)	282.5 g/mol
10.	Hexaflumuron	N-((3,5-Dichloro-4-(1,1,2,2-tetrafluoroethoxy)phenyl)carbamoyl)-2,6-difluorobenzamide	461.14 g/mol
11.	2, 6 Difluorobenzoic acid	Monoaromatic carboxylic acid	158.10 g/mol
12.	Diflunisal	5-(2,4-difluorophenyl)-2-hydroxybenzoic acid	250.19 g/mol
13.	Benzoic acid	Monoaromatic Carboxylic acid	122.12 g/mol
14.	Flupyradifurone	4-((6-chloro-3-pyridylmethyl)(2,2-difluoroethyl)amino)furan-2(5H)-one	288.6 g/mol
15.	Difluoroacetic acid	Monoaliphatic carboxylic acid	96.03 g/mol
16.	Glucose	(3R,4S,5S,6R)-6-(hydroxymethyl)oxane-2,3,4,5-tetrol	180.16 g/mol
17.	D ₂ O, DMSO-d ₆ , Acetone-d ₃	Deuterated Solvents	
18.	Trifluoroethanol	Fluorinated Alcohol (<i>TCI chemicals</i>)	100.04 g/mol
19.	TEMPOL (C ₉ H ₁₈ NO ₂)	4-hydroxy-2,2,6,6-tetramethylpiperidin-1-oxyl	172.24 g/mol
20.	Dipotassium Phosphate	Potassium hydrogenphosphate (K ₂ HPO ₄)	174.2 g/mol
21.	Monopotassium Phosphate	Potassium dihydrogenphosphate (KH ₂ PO ₄)	136.08 g/mol
22.	HEPES Powder for Buffer	N-2-hydroxyethylpiperazine-N-2-ethanesulfonic acid	238.3 g/mol

A2. NMR Pulse Parameters

The general pulse parameters used during the acquisition of NMR spectrum for different nuclei and relevant to the present Thesis are provided in table A2.

Table A2: Values of various pulse parameters kept during acquisition of NMR of different nuclei.

Pulse Parameters	¹⁹ F (470.7 MHz)	¹ H (500 MHz)	² H (76.7 MHz)	¹³ C (125 MHz)
P ₁ (μs) (90° hard pulse width)	15	14.15	390	10.60
PLW1 [W, dbW] (power level: 90° pulse)	19, -12.79	15, -11.76	1.54, -1.88	66, -18.20
P ₂ (μs) (180° hard pulse width)	30	28.30	780	21.20
Decoupling scheme for protons	Waltz16	--	--	Waltz16
PLW2 [W, dbW] (power level: decoupling)	0.469, 3.29	--	--	0.469,3.29 (1 st) 0.236,6.27 (2 nd)
PCPD2 (μs) (90° pulse: decoupling)	80	--	--	80
P18 (μs) width of shaped pulse used for solvent presaturation	--	100000 (Squa10.1000)	--	--
SPW6 [W, dbW] (power level: shaped pulse)		0.00028, 35.48	--	--

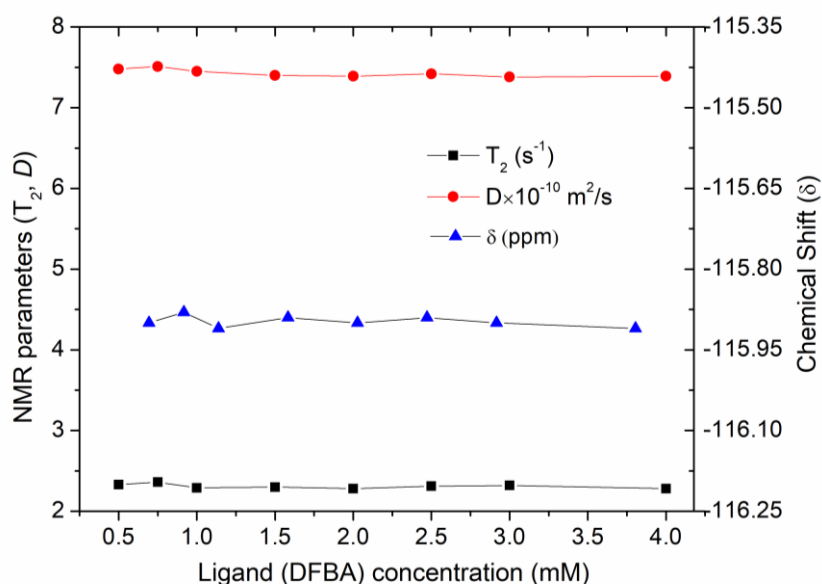


Figure A1: Values of NMR parameters i.e. chemical shift (δ), transverse relaxation time (T_2), Diffusion coefficient (D) as the function of DFBA concentration in absence of BSA. The values of these parameters are found nearly same with increasing DFBA concentration ruling out any possibility of aggregation.

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Annexure B

Section B1. EXTRACTION OF HUMIC ACID: INTERNATIONAL HUMIC SUBSTANCES SOCIETY (IHSS) METHOD

Humic acid is extracted from soil with a procedure recommended and used by the IHSS (International Humic Substances Society) to isolate humic acid. The soil sample is dried and sieved through a 1 cm sieve. Then the sample is treated with 0.1 M HCl at room temperature to have a final ratio of solution to raw sample as 10:1. The suspension is shaken on rotating shaker for 12 h and the supernatant is discarded by centrifugation. The remaining soil residue is neutralized to pH 7.0 using 1 M NaOH and further 0.1 M NaOH is again added to get a final ratio of solution to soil as 10:1. The suspension is extracted by shaking for 24 h in N₂ atmosphere. The alkaline suspension is allowed to settle overnight and the supernatant is collected by centrifugation. The supernatant is acidified with 6 M HCl with constant stirring till pH 1.0 and then allowed to stand for 24 h. It is then centrifuged to separate the humic acid (precipitate) fraction. The humic acid fraction is redissolved by adding a minimum volume of 0.1 M KOH and then solid KCl is added to obtain the final concentration of 0.3 M [K⁺]. Afterwards, it is centrifuged at high speed to remove the suspended solids. The humic acid is reprecipitated by adding 6 M HCl with constant stirring to pH 1.0 and then allowed to stand again for 12 h. The sample is centrifuged and the supernatant discarded. The humic acid precipitate is treated in a mixture of 0.1 M HCl - 0.3 M HF solution in a plastic container and shaken for 12 h at room temperature. Treatment by HCl-HF is repeated once again after discarding the supernatant to obtain sufficiently low ash content. It is centrifuged once again for the last time and the supernatant is discarded. The final portion of humic acid is obtained after washing with distilled water and subsequent drying.

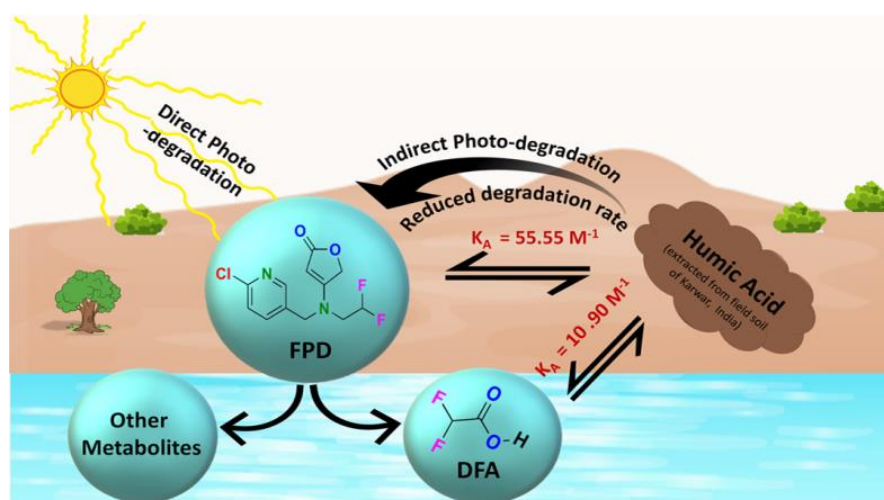


Figure B1: Graphical representation of the Part-II investigation of chapter 4 showing kinetics of direct photo-degradation of FPD in UV light and in presence of KHA forming DFA and other metabolites.

Section B2: Characterization of HA by UV–visible spectroscopic measurements

Figure B2 represents the absorption spectra of extracted (KHA) and Aldrich (AHA) HA that clearly demonstrate similar profile of absorption spectra with a significant difference in absorbance intensity below 350 nm. The spectra are rather featureless, representing only a decrease in absorbance with increasing wavelength indicative of various chromophoric groups present in HA.

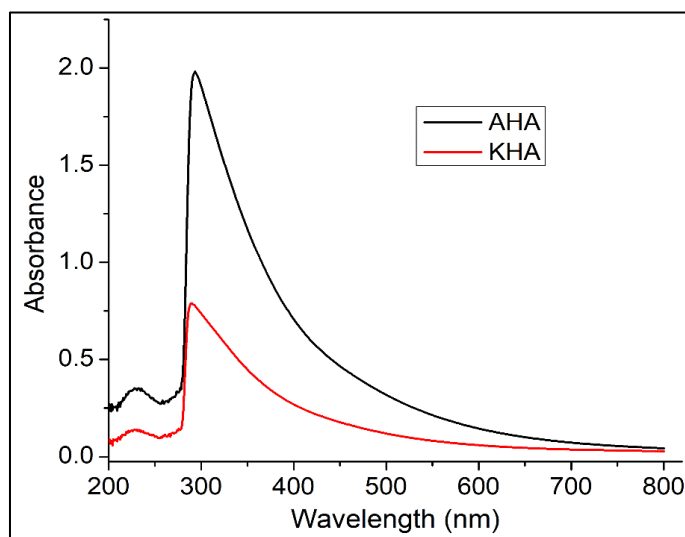


Figure B2: UV–vis absorption spectra of commercial (AHA) and extracted HA (KHA) in aqueous PB buffer (pH=7.4) at 298 K.

Interestingly, it is observed that a minor shoulder is present around 260 nm for both AHA and KHA confirming the presence of aromatic structure in organic HS. It is also well known that this shoulder between 230–270 nm can be due to the conjugation of ketone and quinone [Ghosh and Schnitzer, 1979]. It is customary in the literature to analyze the ratio of absorbance measured at two different wavelengths, e.g., λ_1 and λ_2 and denoted as E_{λ_1/λ_2} (known as optical ratio or color ratio) to analyze the chemical composition of HA. Table B1 documents the important ratios of absorbance values widely used for characterization of HA *viz.*, $A_{280/365}$, $A_{280/665}$, and $A_{465/665}$ measured for both AHA and KHA from UV-vis spectra and denoted respectively as E_2/E_3 , E_2/E_6 and E_4/E_6 . A relatively higher E_4/E_6 and E_2/E_3 ratio is observed for KHA compared to AHA. Such high value accounts for lower humification index, implying a lower degree of condensation of the aromatic carbon chain of HA [Morán Vieyra, et al., 2009]. Higher E_4/E_6 ratio also indicates presence of relatively higher aliphatic components in KHA compared to AHA. The E_4/E_6 ratio is proportional to the acidity and can be inversely related to molecular weight, particle size, and the degree of aromaticity [Kang, et al., 2002]. On the other hand, the lower E_2/E_6 represents the increasing humification of organic components from the early transformation stage, like lignins and quinones for HA [Uyguner, et al., 2004]. These values are in general different for different HA and solely depends on the soil from which the particular HA is extracted. The value of the $\Delta \log K$ coefficient [Sim et al., 2006] ($\Delta \log K = \log A_{400} - \log A_{600}$), known as Kumada Parameter, has been calculated to get additional proof on the less degree of humification of KHA than AHA [Boguta and Soko, 2016]. Kumada categorizes HA as medium to weakly humified (KHA here) if $\Delta \log K$ oscillated from 0.7 to 0.93, while highly humified (AHA in present case) when value is less than 0.7.

Table B1: Value of UV-vis absorbance at various wavelengths for HA in aqueous PB (pH=7.40) and respective optical ratio recorded at 298 K.

HA	A ₂₈₀	A ₃₅₀	A ₄₆₅	A ₆₆₅	A ₄₀₀	A ₆₀₀	A ₂₈₀ /A ₃₅₀ (E _{2/3})	A ₄₆₅ /A ₆₆₅ (E _{4/6})	A ₂₈₀ /A ₆₆₅ (E _{2/6})	ΔlogK
(AHA) Commercial	0.411	1.171	0.417	0.091	0.707	0.145	0.35	4.58	4.52	0.68
(KHA) Extracted	0.315	0.666	0.239	0.040	0.401	0.075	0.47	5.97	7.88	0.73

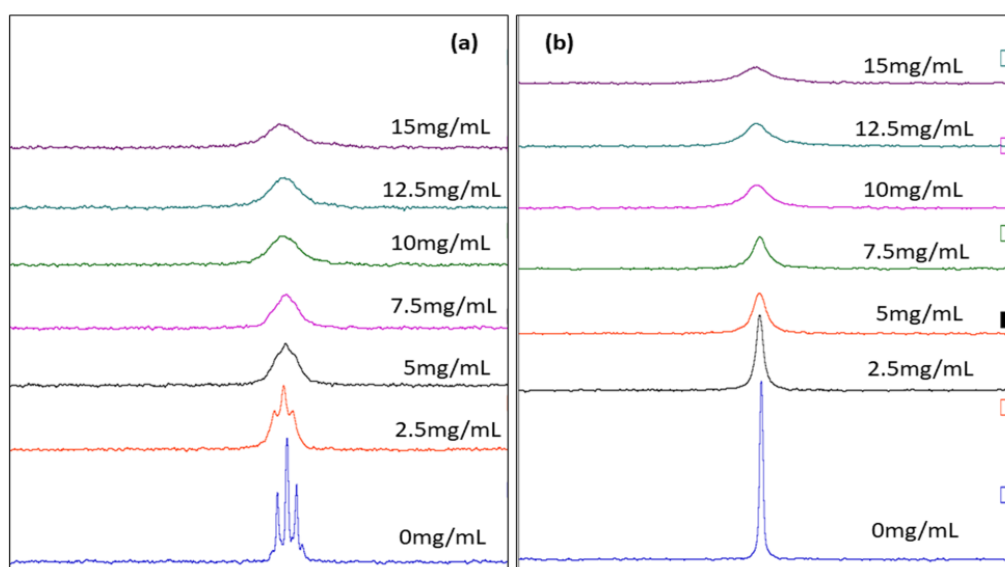


Figure B3: (a) ¹H coupled ¹⁹F NMR (b) ¹H decoupled ¹⁹F NMR spectrum of DFBA as the increasing function of AHA concentration recorded at pH=7.4 and T=298 K.

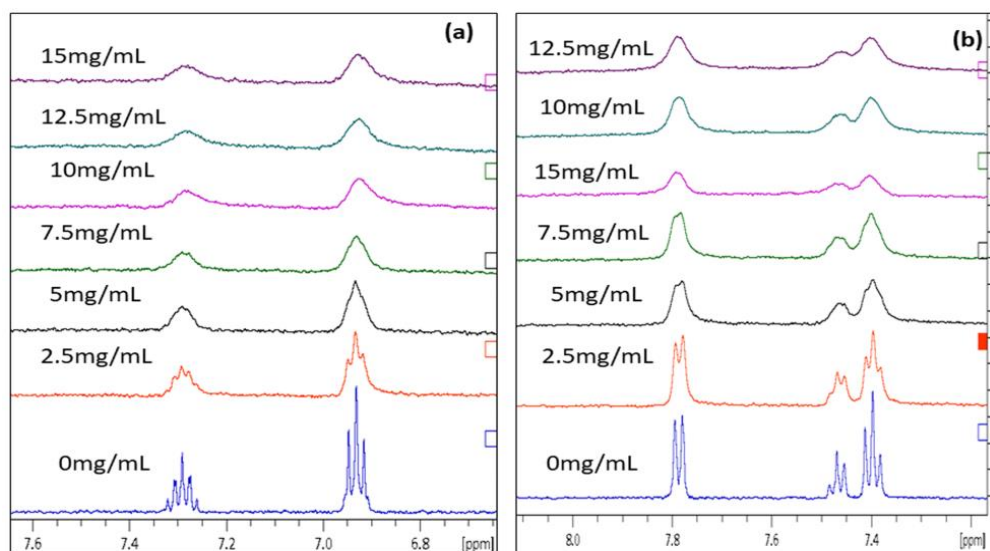


Figure B4: ¹H NMR spectrum of (a) DFBA and (b) BA as the increasing function of AHA concentration at pH=7.4 and T=298 K.

Table B2: T_1 and T_2 relaxation and correlation times (τ_c) for DFBA (2 mM) as a function of the AHA concentrations (mg/mL) at 298 K and pH 7.4. [Those values of T_1 and T_2 are reported where fitting parameter Pearson R^2 correlation coefficient found nearly equal to 1 (while extraction relaxation time from the fitted plot)]

Sample	T_1 (s)			T_2 (s)			Correlation Time τ_c (ns)	
	H_a	H_b	^{19}F	H_a	H_b	^{19}F	H_a	H_b
DFBA								
0 mg/mL HA	8.19	7.16	2.88	5.78	5.15	0.151	0.0133 (determined from ^{13}C T_1)	
2.5 mg/mL HA	5.04	4.75	2.69	0.093	0.077	0.075	2.53	2.70
5 mg/mL HA	4.11	3.8	2.53	0.054	0.044	0.060	3.01	3.22
7.5 mg/mL HA	3.50	3.29	2.45	0.042	0.036	0.038	3.16	3.32
10 mg/mL HA	2.61	2.51	2.28	0.031	0.026	0.033	3.18	3.42
12.5 mg/mL HA	2.29	2.25	2.21	0.026	0.022	0.024	3.25	3.53
15 mg/mL HA	2.03	1.96	2.17	0.023	0.018	0.018	3.26	3.65
25 mg/mL HA	1.40	1.43	1.83	0.015	0.013	0.151	3.36	3.67

Table B3: T_1 and T_2 relaxation and correlation (τ_c) times for BA (2 mM) as a function of the AHA concentrations (mg/mL) at 298 K and pH 7.4.

Sample	T_1 (s)			T_2 (s)			Correlation Time τ_c (ns)		
	H_a	H_c	H_b	H_a	H_c	H_b	H_a	H_c	H_b
BA									
0 mg/mL HA	5.56	6.15	6.85	4.14	4.17	4.24	0.0121 (determined from ^{13}C T_1)		
2.5 mg/mL HA	4.90	5.02	5.36	0.143	0.140	0.138	2.08	2.12	2.19
5 mg/mL HA	3.94	4.15	4.36	0.087	0.086	0.085	2.34	2.41	2.47
7.5 mg/mL HA	2.81	3.00	3.13	0.051	0.047	0.049	2.56	2.75	2.75
10 mg/mL HA	2.35	2.24	2.40	0.034	0.032	0.034	2.87	2.88	2.89
12.5 mg/mL HA	2.09	1.88	2.00	0.029	0.026	0.028	2.93	2.93	2.91
15 mg/mL HA	1.81	1.74	1.75	0.025	0.024	0.024	2.93	2.93	2.95
25 mg/mL HA	----	0.93	0.91	----	0.011	0.011	----	3.18	3.14

Table B4: Observed (Raw) and viscosity corrected ^{19}F & ^1H diffusion coefficient (D) Values ($10^{-10} \text{ m}^2 \text{ s}^{-1}$) for signals of 2 mM DFBA & BA progressively added with AHA (mg mL $^{-1}$) at 298 K and pH 7.4.

Sample	HA (mg/mL)	Measurement for DFBA				Measurement of BA			
		$\eta[\text{sol}]/\eta[0]$	Average $^1\text{H}_{\text{DFBA}}$ (raw)	Average $^1\text{H}_{\text{DFBA}}$ (corrected)	$^{19}\text{F}_{\text{DFBA}}$ (raw)	$^{19}\text{F}_{\text{DFBA}}$ (corrected)	$\eta[\text{sol}]/\eta[0]$	Average $^1\text{H}_{\text{BA}}$ (raw)	Average $^1\text{H}_{\text{BA}}$ (corrected)
	0	1.00	5.875	5.875	6.093	6.093	1.000	6.397	6.397
	2.5	1.005	5.556	5.587	5.852	5.88	1.020	6.057	6.178
	5	1.022	4.744	4.851	5.561	5.68	1.089	5.428	5.911
	7.5	1.037	4.157	4.314	5.305	5.505	1.133	4.828	5.470
	10.0	1.053	3.842	4.049	5.092	5.366	1.189	4.234	5.034
	12.5	1.148	3.496	4.015	4.734	5.437	1.121	3.876	4.689
	15.0	1.225	3.243	3.973	4.424	5.420	1.251	3.606	4.511
	25	1.286	3.045	3.915	4.211	5.414	1.321	2.911	3.846

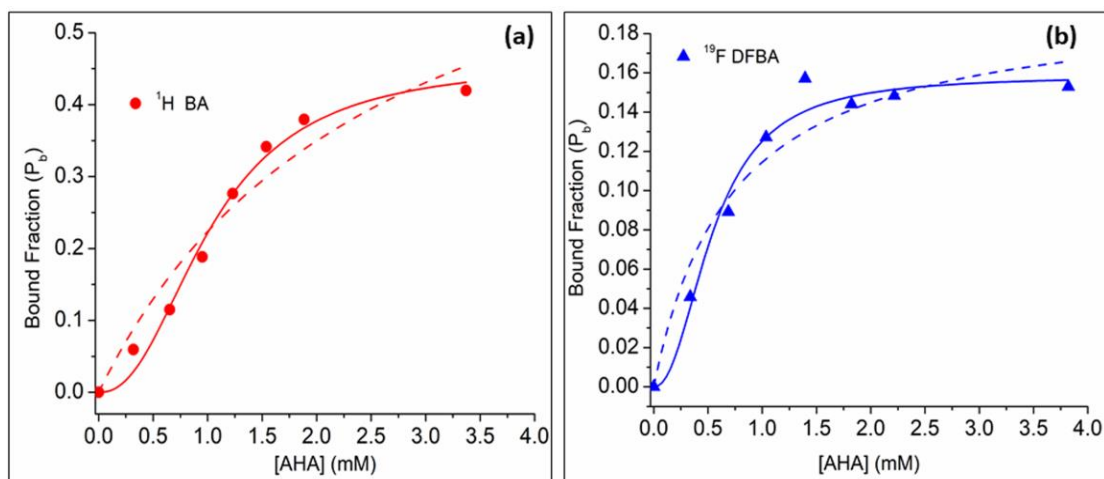


Figure B5: Plot for K_A determination of (a) BA and (b) DFBA using P_b values calculated from $^1\text{H } D_{\text{BA}}$ and $^{19}\text{F } D_{\text{DFBA}}$ values, respectively. (solid line and dash line represents the fitting obtained by modified Langmuir model and Langmuir model respectively)

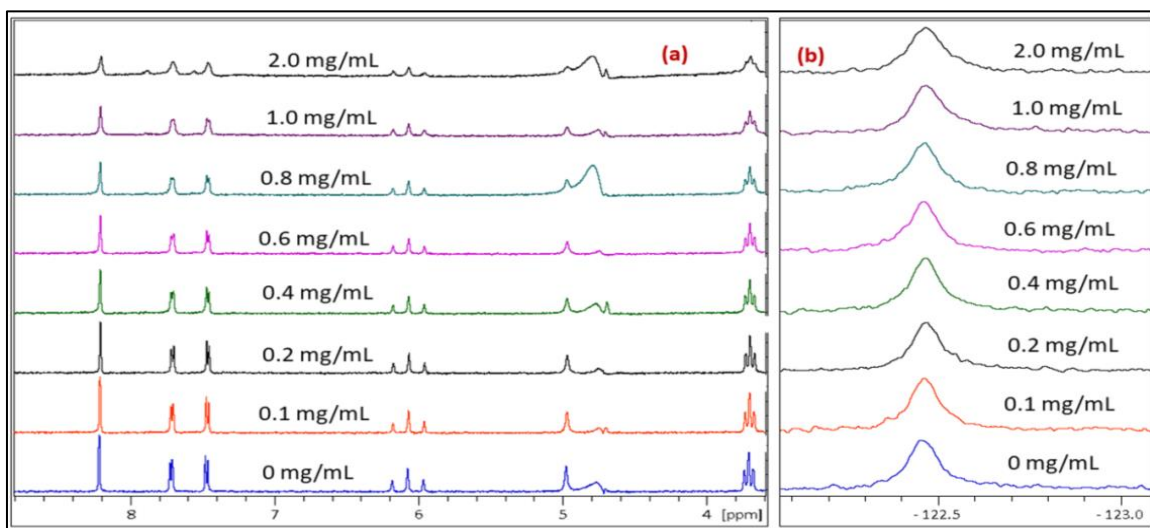


Figure B6: (a) Solvent presaturated ^1H NMR (b) ^1H decoupled ^{19}F NMR spectrum for FPD as a function of KHA concentration prepared in 5: 95 DMSO: aqueous PB solvent system recorded at $T=295\text{ K}$ and $\text{pH}=7.40$.

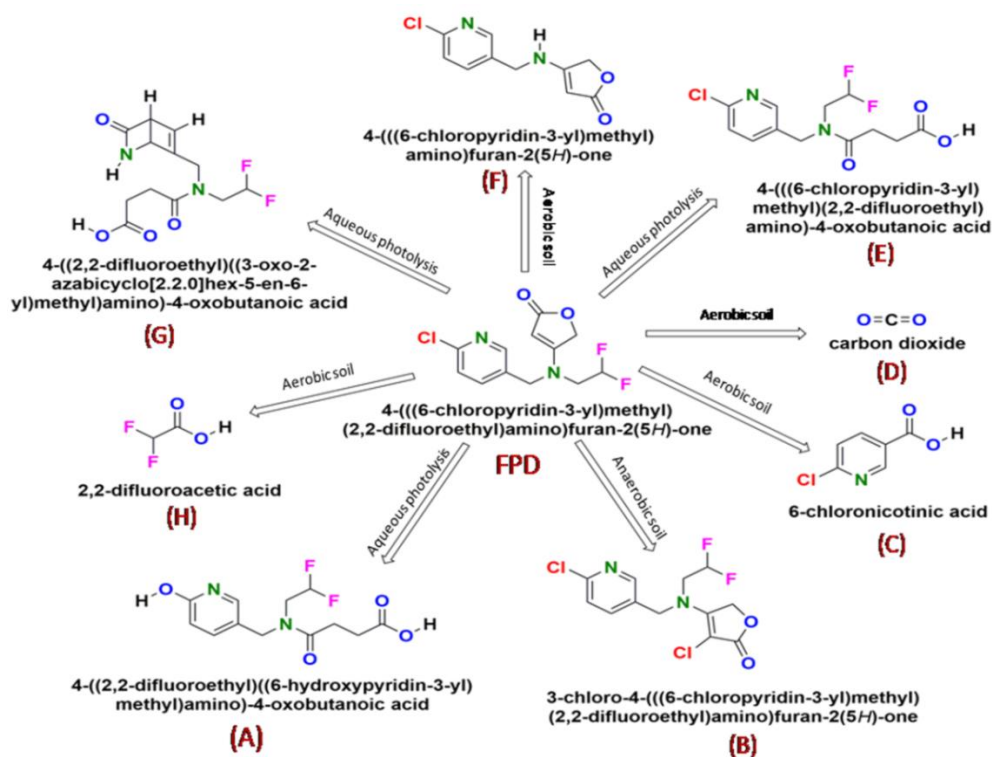


Figure B7: Different possible degradation products for FPD from various degradation pathways. [Source : (Flupyradifurone, 2013; Glaberman and Katrina, 2014)]

Section B3: For free FPD, the plausible mechanism for degradation can be anticipated as: the generation of reactive $\cdot\text{OH}$ (hydroxyl) radical and electron from OH^- (due to the interaction of UV with aqueous PB) are known to oxidize the organic compounds (FPD in the present case). Environmental implications of ($\cdot\text{OH}$) are well known as they are responsible for the formation of secondary pollutants by inducing the chemical transformation of a number of primary pollutants. Also, they bears the second highest oxidation potential (2.80 V), after the strongest oxidant $-\text{F}$. The electrophilic nature (oxidant species) of the $\cdot\text{OH}$ has the potential of oxidizing almost all electron rich organic moieties non-selectively; first generating degraded metabolites and further eventually converting them to CO_2 and H_2O [Si, et al., 2004].

Table B5: ^1H and ^{19}F NMR T_1/T_2 relaxation ratio for FPD and DFA as an increasing function of KHA prepared in 5: 95 DMSO: aqueous PB solvent (pH=7.40) and measured at 295 K.

Sample	T_1/T_2 ratio for FPD						T_1/T_2 ratio for DFA	
	H_a	H_b	H_c	H_g	H_h	^{19}F	H_a	^{19}F
0 mg/mL HA	8.36	4.79	5.93	6.81	2.64	50.58	1.80	1.31
0.1 mg/mL KHA	11.09	6.74	7.52	7.36	2.98	52.05	2.01	1.50
0.2 mg/mL KHA	12.18	8.19	8.46	7.64	3.35	52.98	2.21	1.59
0.4 mg/mL KHA	12.98	10.0	9.09	7.78	3.74	54.28	2.43	1.69
0.6 mg/mL KHA	13.77	11.05	9.45	7.79	4.00	55.86	2.62	1.88
0.8 mg/mL KHA	14.90	11.20	9.81	7.86	4.30	57.01	2.74	2.02
1 mg/mL KHA	15.65	11.30	10.2	7.92	4.57	59.73	2.87	2.15
2 mg/mL KHA	16.71	11.79	11.32	8.045	4.82	61.00	3.03	2.59

Table B6: Representative self-diffusion coefficient (D) values measured for 1 mM FPD (aromatic protons) and DFA as a function of KHA concentration in 5: 95 DMSO: aqueous PB solvent (pH=7.40) at 295 K.

Sample FPD / DFA (1mM)	Viscosity Corrected $D \times 10^{-10} \text{ m}^2/\text{s}$ (FPD)			Fraction of bound FPD (P_b) (%)	Viscosity Corrected $D \times 10^{-10} \text{ m}^2/\text{s}$ (DFA)		Fraction of bound DFA (P_b) (%)
	H_a	H_b	H_c		H_a	^{19}F	
0 mg/mL HA	2.80	2.99	2.76	0	8.72	9.25	0
0.1 mg/mL HA	2.72	2.86	2.61	8.7	8.4	8.89	4.8
0.2 mg/mL HA	2.67	2.74	2.51	16.2	8.12	8.42	8.9
0.4 mg/mL HA	2.55	2.53	2.45	30.2	7.88	8.05	12.5
0.6 mg/mL HA	2.39	2.39	2.35	39.6	7.56	7.79	17.2
0.8 mg/mL HA	2.24	2.26	2.22	51.4	7.24	7.56	22.0
1 mg/mL HA	2.00	2.07	1.99	62.8	6.97	7.24	26.1
2 mg/mL HA	1.75	1.80	1.40	78.5	6.55	6.79	32.3

Section B4: Figure B8 (I and II) represents the ^1H - ^1H STD and ^{19}F - ^1H STD experiments respectively recorded for DFA-KHA system. Peaks of significant intensity appeared in both ^1H - ^1H STD and ^{19}F - ^1H STD spectrum that confirms that -F of DFA is taking part while interacting with HA. GEM is not possible for DFA due to presence of single peak in ^1H and ^{19}F NMR spectra. This observation supports the ^1H and ^{19}F chemical shift and line-broadening experiments of DFA shown in figure B9 (a & b). ^1H spectra of DFA in presence of 1 mg/mL KHA exhibited a maximum downfield shift by 2.75Hz only while ^{19}F shifted upfield of maximum 15.93 Hz. As per literature, the formation of H-bond between -F and HA or solvent molecule increases the electron density around fluorine atoms, thus causing upfield shift for F nuclei due to enhanced shielding [Šmejkalová et al., 2009]. The changes in ^{19}F chemical shift for DFA also gives a clue of involvement of -F moiety while binding with HA, unlike FPD where least change is seen in chemical shift of ^{19}F and proton near to -F. The variable drift in chemical shift DFA in presence of HA compared to FPD is surely evident for the differential binding modes of both the molecules with HA as evident from STD experiments as well.

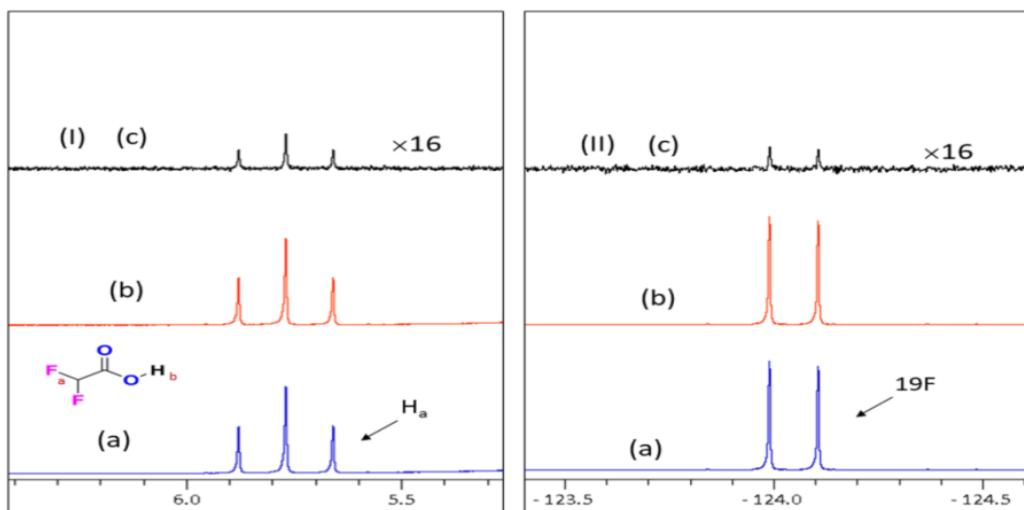


Figure B8: (I) ^1H - ^1H STD NMR (II) ^{19}F - ^1H STD NMR (a) STD_{off} (b) STD_{on} and (c) STD_{diff} spectrum for DFA-HA system in 5: 95 DMSO: aqueous PB solvent (pH=7.4) at $T = 295$ K.

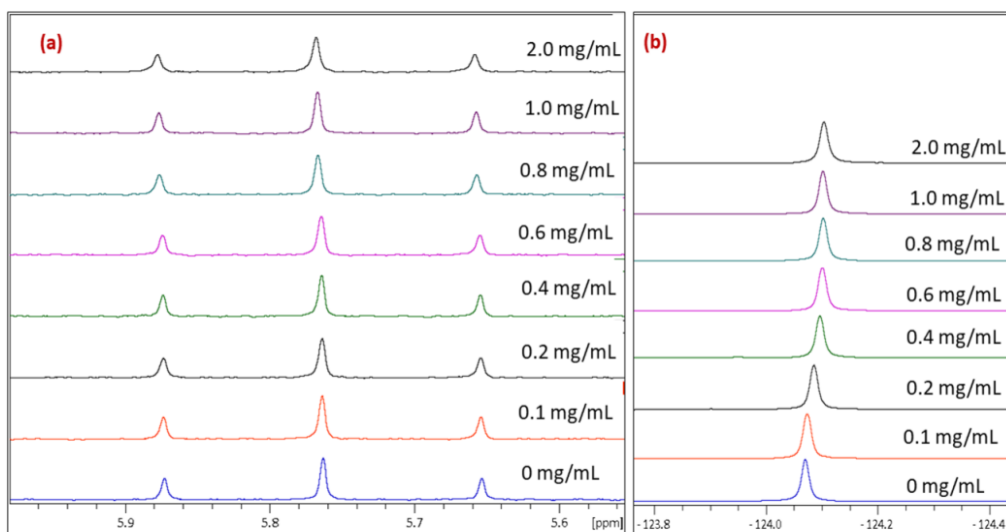


Figure B9: Solvent suppressed ^1H NMR (b) ^1H decoupled ^{19}F NMR spectrum for DFA as a function of HA concentration in 5: 95 DMSO: aqueous PB solvent (pH=7.4) at $T = 295$ K.

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- 2) **Chaubey, B.**, Narawal, P., Khandelwal, A. & Pal, S. Aqueous photo-degradation of Flupyradifurone (FPD) in presence of a natural Humic Acid (HA): A quantitative solution state NMR analysis. *Photochemistry and Photobiology A: Chemistry*, **2020**, 405 (2021), 2020, 112986.
- 3) **Chaubey, B.**, Dey, A., Banerjee, A., Chandrakumar, N. & Pal, S. Assessment of TFE solvent dynamics inducing conformational transitions in Melittin: An approach with solvent ^{19}F low field NMR relaxation and Overhauser DNP studies. *Journal of Physical Chemistry B.*, 124 (28), **2020**, 5993–6003.
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Peer reviewed International Journals (Under preparation)

- 1) **Chaubey, B.**, Chandrakumar, N. & Pal, S. Preferential solvation of carbohydrates in water-trifluoroethanol mixtures: A solvent detected heteronuclear NMR approach. To be submitted in *Physical Chemistry Chemical Physics*, **2021**.

Peer reviewed International Conference proceedings (Published)

- 1) **Chaubey, B.**, & Pal, S. Hexaflumuron- β -Cyclodextrin Inclusion Complex as Improved Pesticide Formulation: A NMR Case Study. *AIP Conference Proceedings*. 2142, **2019**, 180003(1)–180003(4).

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- 1) Vandana, **Chaubey, B.**, Dhaharwal, A. K., & Pal, S. Solvent-dependent binding interactions of the organophosphate pesticide, chlorpyrifos (CPF), and its metabolite, 3,5,6- trichloro-2-pyridinol (TCPy), with Bovine Serum Albumin (BSA): A comparative fluorescence quenching analysis. *Pesticide Biochemistry and Physiology*, 139, **2017**, 92-100.
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- 4) **Chaubey, B.** & Pal, S. (Oral) "Quantitative Investigation of Interaction between Aqueous Humic Acid and 2, 6 Difluorobenzoic Acid by ^{19}F & ^1H NMR Spectroscopy' at Conference on Magnetic Resonance in Medicine & 25th National Magnetic Resonance Society Meeting (NMRS-2019) organized by AIIMS and INSA, New Delhi.
- 5) **Chaubey, B.** & Pal, S. (Oral) 'Structural transition of aqueous Melittin in presence of Trifluoroethanol: A NMR relaxometry approach" at 3rd North-West Meeting on Spectroscopy, Structure and Dynamics organized by IIT Roorkee, **2019**.
- 6) **Chaubey, B.** & Pal, S. (Poster) 'Hexaflumuron- β -Cyclodextrin Inclusion Complex as Improved Pesticide Formulation: A NMR Case Study" at International Conference on Advances in Basic Sciences **2019** (AIP conference proceedings) at Bahal, India.
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- 8) **Chaubey, B.**, Participated in "22nd Conference of National Magnetic Resonance Society of India (NMRS-2016)" at Department of Chemistry, Indian Institute of Technology Kharagpur.

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